# Gene sets & correlation

Michael Love June 8, 2015 Gene set tests with correlation (from PH525x notes and Barry, Nobel and Wright, 2008)

Consider the average *t*-statistic from *N*|genes in a set *G*|:

$$\bar{t} = \frac{1}{N} \sum_{i \in G} t_i$$

This statistic  $\overline{t}$  combines the information about DE from the set and might be a useful test statistic.

#### Gene set tests with correlation

Under the null hypothesis, the *t* have mean 0. If the *t* are independent then  $\sqrt{Nt}$  has standard deviation 1 and is approximately normal:

 $\sqrt{N}\bar{t} \sim N(0,1)$ 

This comes from the following decomposition of the variance:

$$\operatorname{Var}(\overline{t}) = \frac{1}{N^2} \operatorname{Var}(t_1 + \dots + t_N)$$
$$= \frac{1}{N^2} \left( \operatorname{Var}(t_1) + \dots + \operatorname{Var}(t_N) \right)$$
$$= \frac{1}{N}$$

#### Gene set tests with correlation

Now consider the case that the test statistics  $t_i$  in a gene set are not independent but have correlation  $\rho_i$  under the null hypothesis.

$$\bar{t} = \frac{1}{N} \sum_{i \in G} t_i$$

$$\operatorname{corr}(t_i, t_{i'}) = \rho, \quad i, i' \in G$$

The variance of the average *t*-statistics will be:

$$\operatorname{Var}(\bar{t}) = \frac{1}{N^2} \operatorname{Var}((1 \dots 1)(t_1 \dots t_N)')$$
  
=  $\frac{1}{N^2} (1 \dots 1) \begin{pmatrix} 1 & \rho & \dots & \rho & \rho \\ \rho & 1 & \rho & \dots & \rho \\ \dots & \dots & \dots & \dots & \dots \\ \rho & \rho & \dots & \rho & 1 \end{pmatrix} (1 \dots 1)'$   
=  $\frac{1}{N^2} \{N + (N - 1)N\rho\}$   
=  $\frac{1}{N} \{1 + (N - 1)\rho\}$ 

# Variance inflation with correlation

So the variance inflation factor (VIF) comparing the independent case to the case with correlation is:

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VIF = 1 + (N - 1)\bar{\rho}|
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So the increased width (standard deviation) of the null distribution for a gene set with 20 genes and average correlation 0.1 will be:

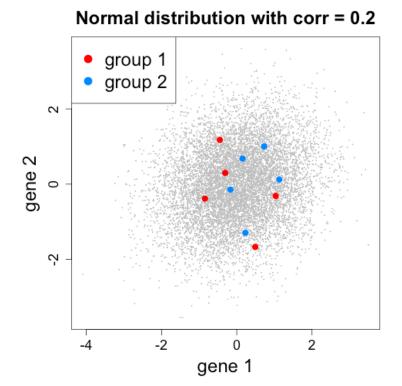
sqrt(1 + 19 \* 0.1)

[1] 1.702939

This VIF is approximately true also for testing the set statistics against the complement: the genes not in the set (see Barry, Nobel and Wright 2008).

# Test statistic vs expression correlations

Here, the expression of 5 samples vs 5 samples, no difference in expression across group but a correlation of gene expression.



Test statistic vs expression correlations If the test statistic T is a linear form of the data X (e.g. log fold change), then:

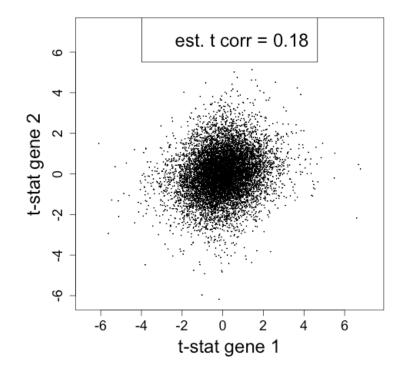
$$\rho_{i,i'}^T = \rho_{i,i'}^X \Big|$$

For t-test, the relationship is monotone, approximately linear and:

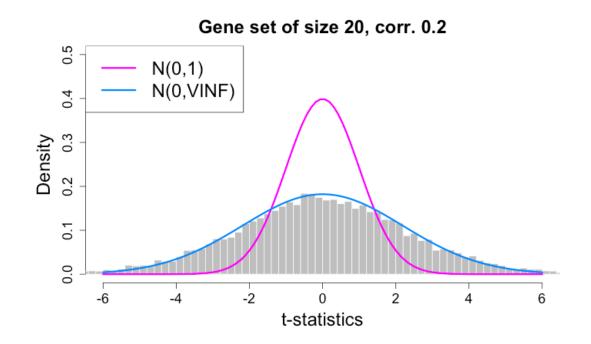
$$\rho_{i,i'}^T \approx \rho_{i,i'}^X \Big|$$

(Barry, Nobel and Wright, 2008)

### Simulate expression correlation of 0.2

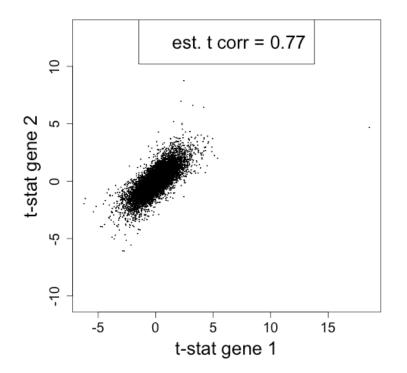


#### Distribution of t-statistics

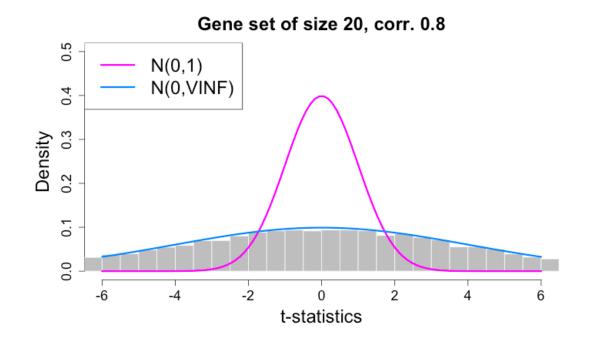


28% of the simulated t-statistics are outside of the center 99% of the N(0, 1) distribution.

## Again, with correlation of 0.8



# Distribution of t-statistics (corr = 0.8)



54% of the simulated t-statistics are outside of the center 99% of the N(0, 1) distribution.

# Intuition

- Suppose expression is correlated within a gene set under the null
- By chance, for some gene, the expression could be high for the group 2 samples, and low for group 1 samples
- The t-statistic will be large and positive for this gene
- Because expression is correlated across genes in the set, other genes will likely see the same pattern
- t-statistics will be correlated within the set

# Why would we see null correlations?

Where do expression correlations *under the null* come from? My guesses in order of importance:

- uncorrected batch effects
- large scale amplifications in cancer
- gene regulatory networks

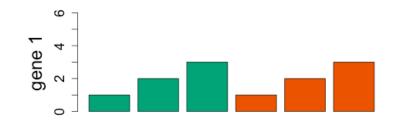
# CAMERA (Wu and Smyth 2012)

for Correlation Adjusted MEan RAnk gene set test, available in the limma package.

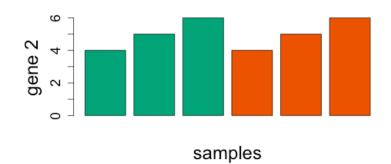
- estimating the inter-gene correlation from the data
- using it to adjust the gene set test statistic
- suitable for any experiment that can be represented by genewise linear models

## Permutations

Assume the null: no differences across condition, although genegene correlation are present



samples

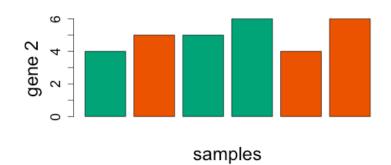


## Permutations

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samples



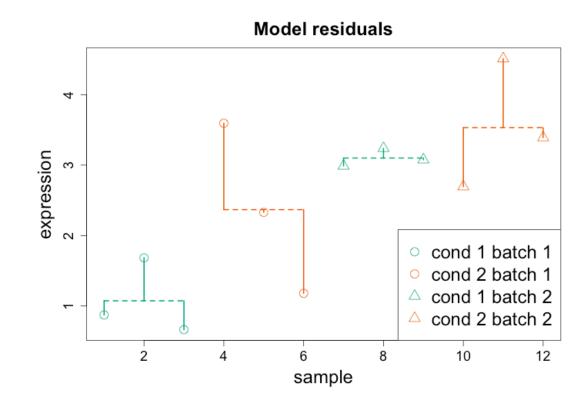
# Permutations

Has limitations:

- only if samples are exchangeable (no batch effects)
- not easy to implement for complex designs (although see SAMseq in the **samr** package for strategies)
- requires sufficient samples for small p-values (although see Larsen and Owen for moment-based trick)

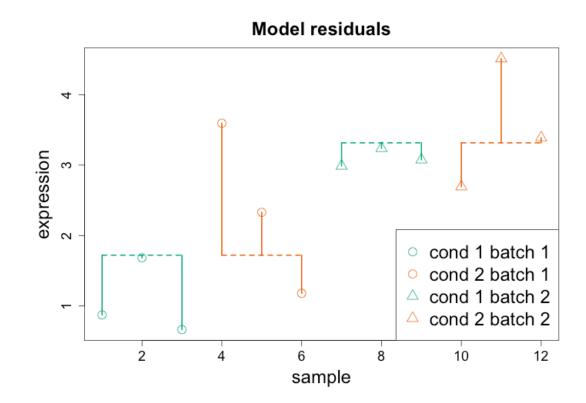
#### Approach using residuals

Suppose we have a 2 condition experiment with 2 batches:



## Approach using residuals

Remove design matrix columns not involving the null hypothesis:



# ROAST (Wu et al. 2010)

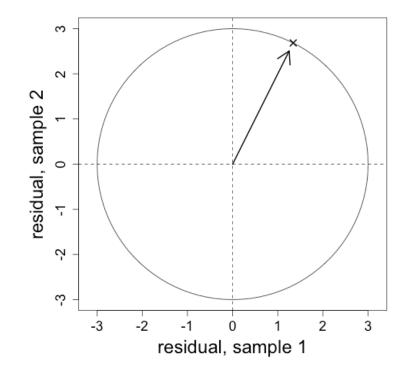
The ROAST method available in the limma package:

Under the null hypothesis (and assuming a linear model) the residuals are independent and identically distributed  $N(0, \sigma_g^2)$ .

We can *rotate* the residual vector for each gene in a gene set, such that gene-gene expression correlations are preserved.

#### What does residual rotation look like?

Like this diagram but around an n-sphere (n, the number of samples).



# ROAST (Wu et al. 2010)

Repeat 10,000 times:

- 1. rotate the residual vector from each gene in the set using the same rotation
- 2. create new data, preserving the gene-gene correlations
- 3. compute test statistics for the rotated data for each gene and compute the gene set statistic

Lastly compare the original gene set statistic to the null distribution from 1-3.

Pros: fast and efficient, fits with any linear model

# Summary

- 1. Gene-gene correlations inflate the null distribution of gene set statistics
- 2. This inflation factor can be directly calculated from the data
- 3. Rotating residuals can also be used to generate a null which incoroporates correlations